

# Molecular Dynamics Analysis

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and Christian Schwantes*

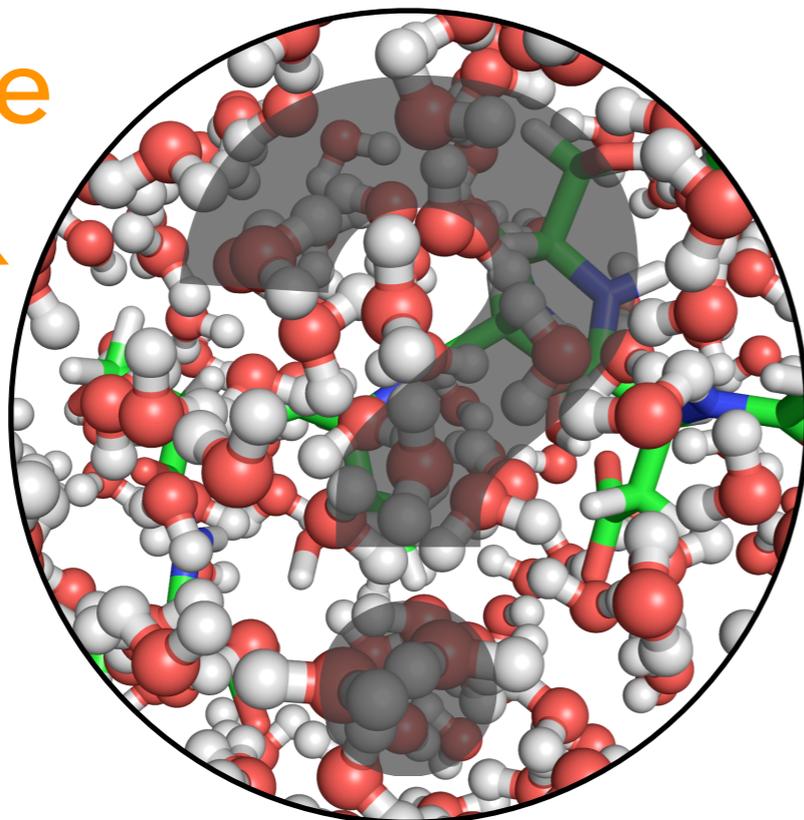
March 29, 2014

# Value of Simulation

- Experiments provide projections of the high-dimensional protein folding process
- Determining the microscopic mechanism from these projections is difficult

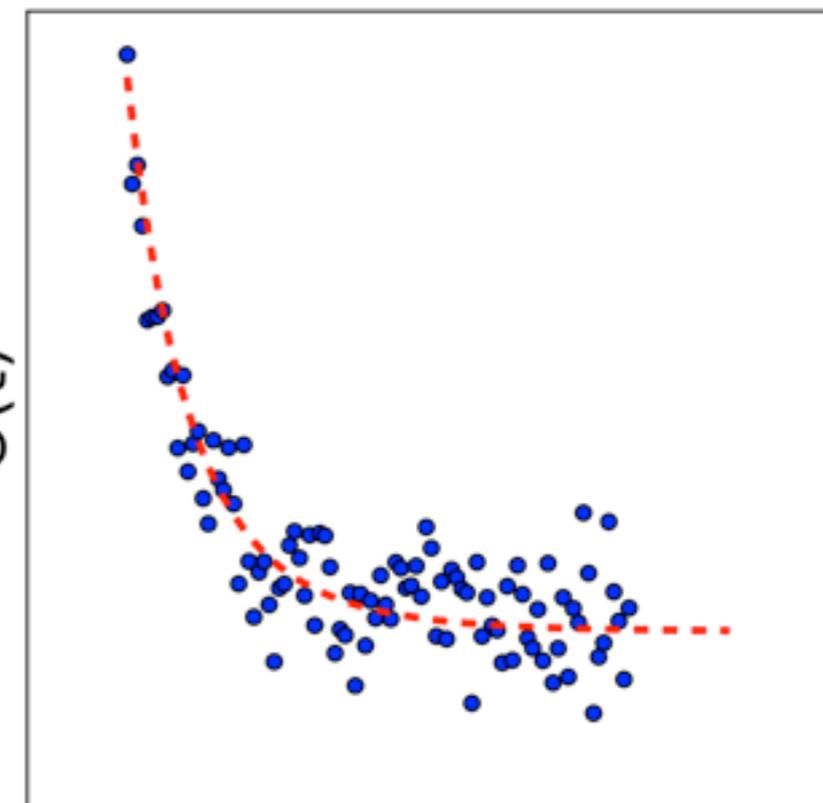
Experimental

Probe



Observable

$O(t)$



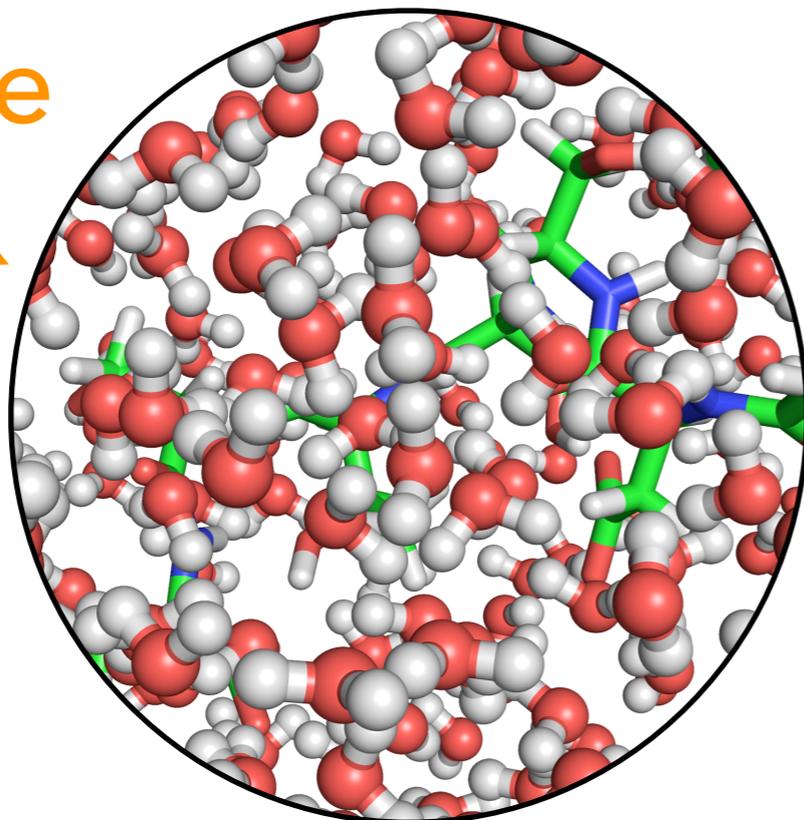
time

# Value of Simulation

- Simulation can provide an atomic-level description that most experiments cannot
- By predicting experimental observables, we can validate our models

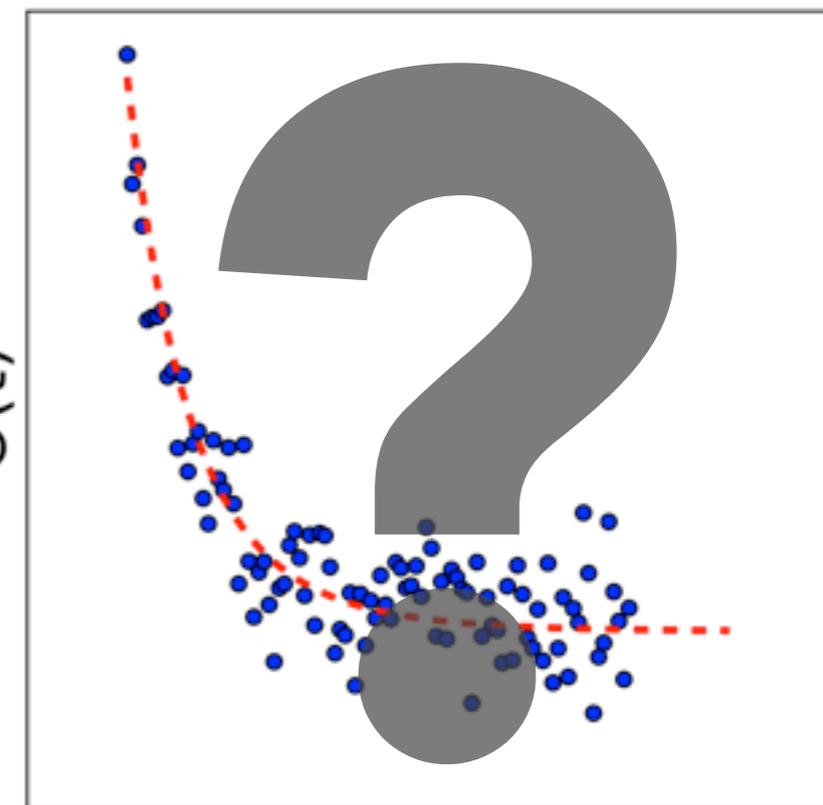
Experimental

Probe



Observable

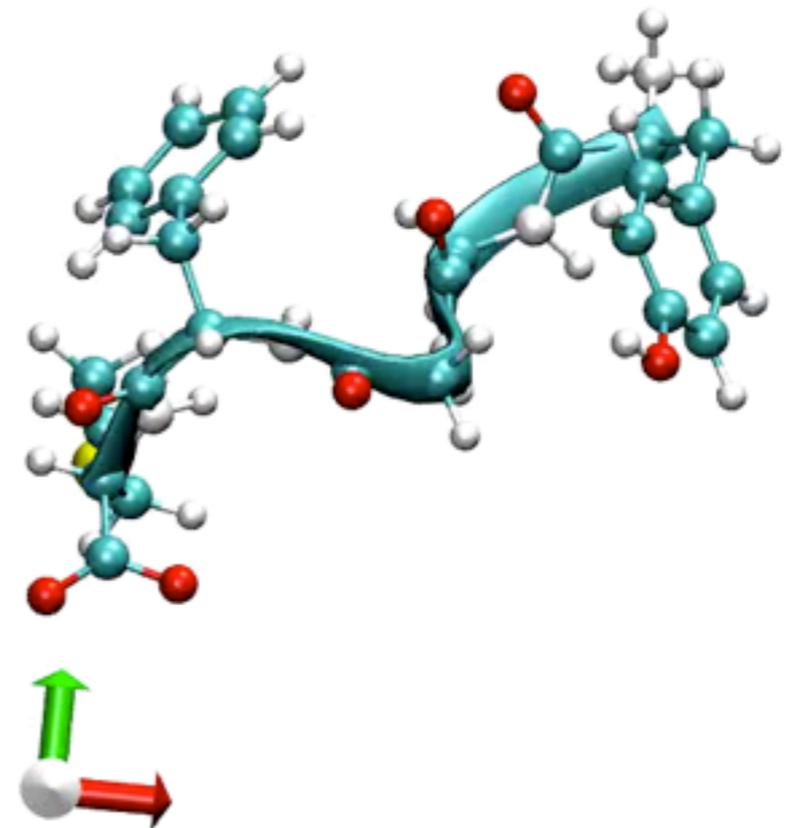
$O(t)$



time

# Molecular Dynamics (MD)

- Let's say we've taken a lot of computer (and human) time to generate a large set of MD trajectories
- ***Now what?***
  - We can certainly make a pretty cool movie
  - But MD is so much more than a YouTube clip!
- We want to understand our results



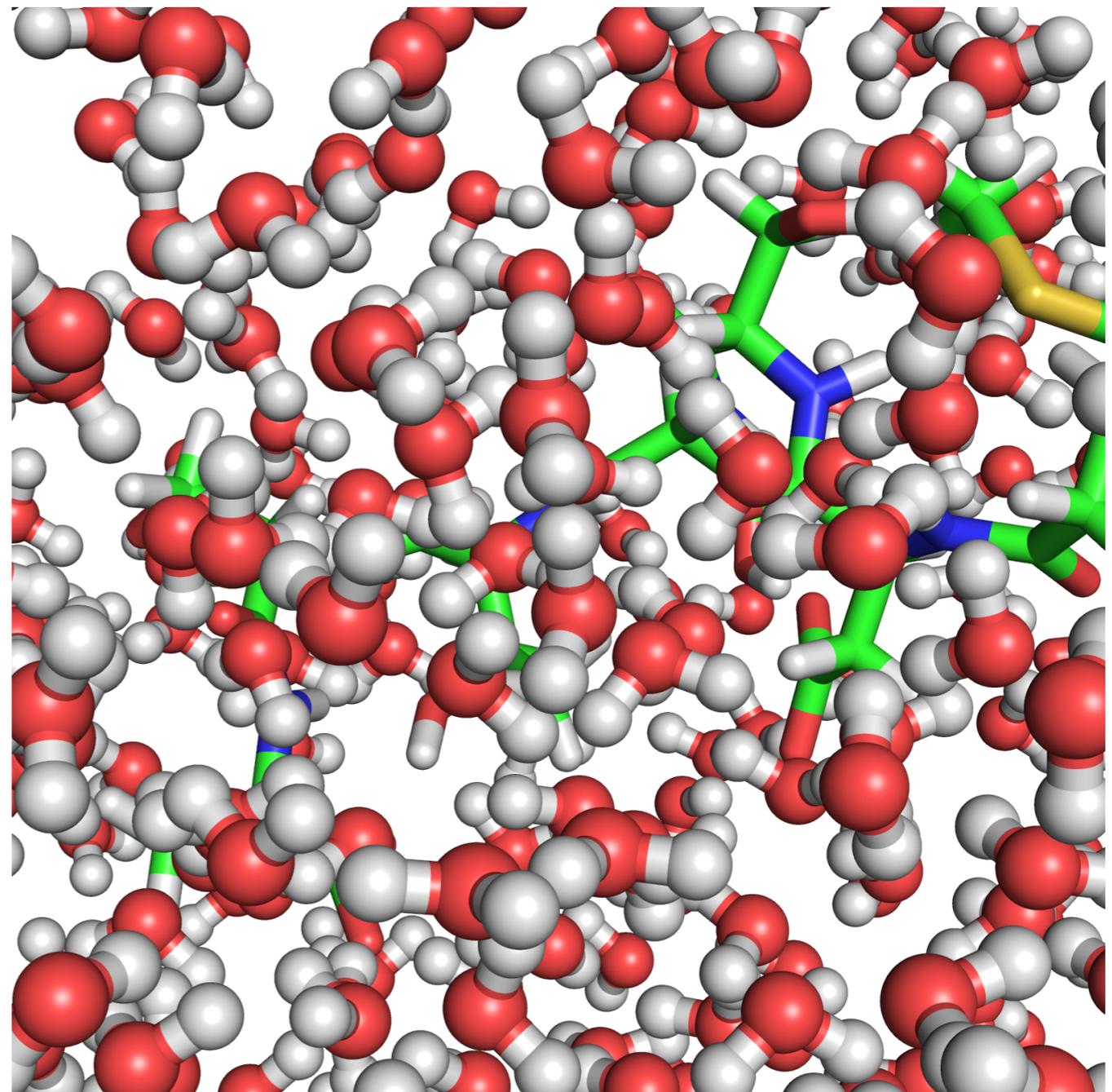
# Quantitative Analysis

- MD datasets are too high-dimensional to simply make sense of out of the box
- A typical molecular dynamics data set has 25,000+ atoms
- We frequently have datasets that are hundreds of microseconds or even milliseconds (millions of frames)
- So we need to simplify!
- But we also want to be sure that we don't simplify in such a way that we lose important information

# Dimensionality Reduction

- We need to simplify the picture in order to make sense of it!
- We already do this!
- Throw out velocities

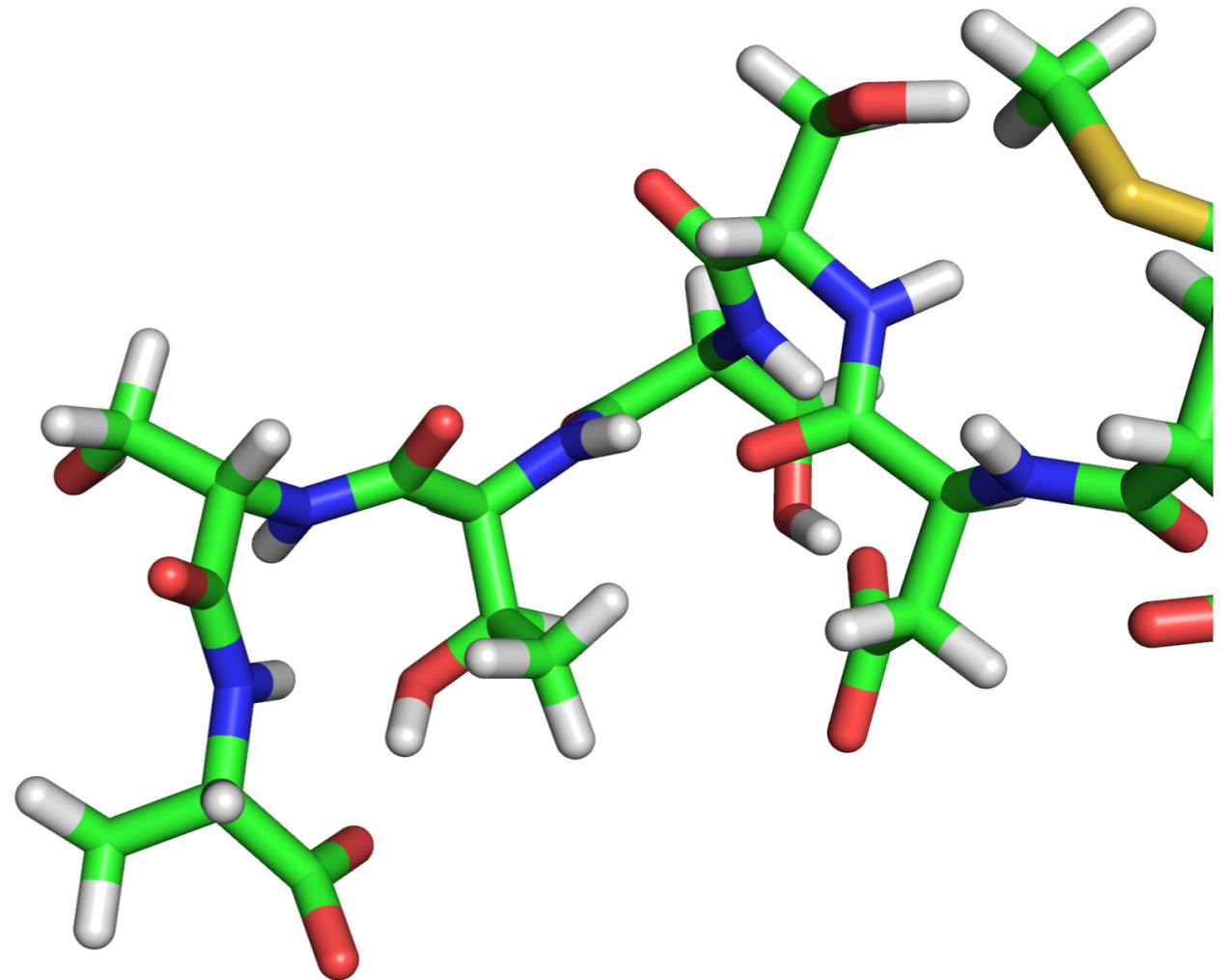
$$(\vec{x}, \vec{v}) \rightarrow (\vec{x}, \cancel{\vec{v}})$$



# Dimensionality Reduction

- We need to simplify the picture in order to make sense of it!
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  - Throw out velocities
  - Throw out solvent degrees of freedom

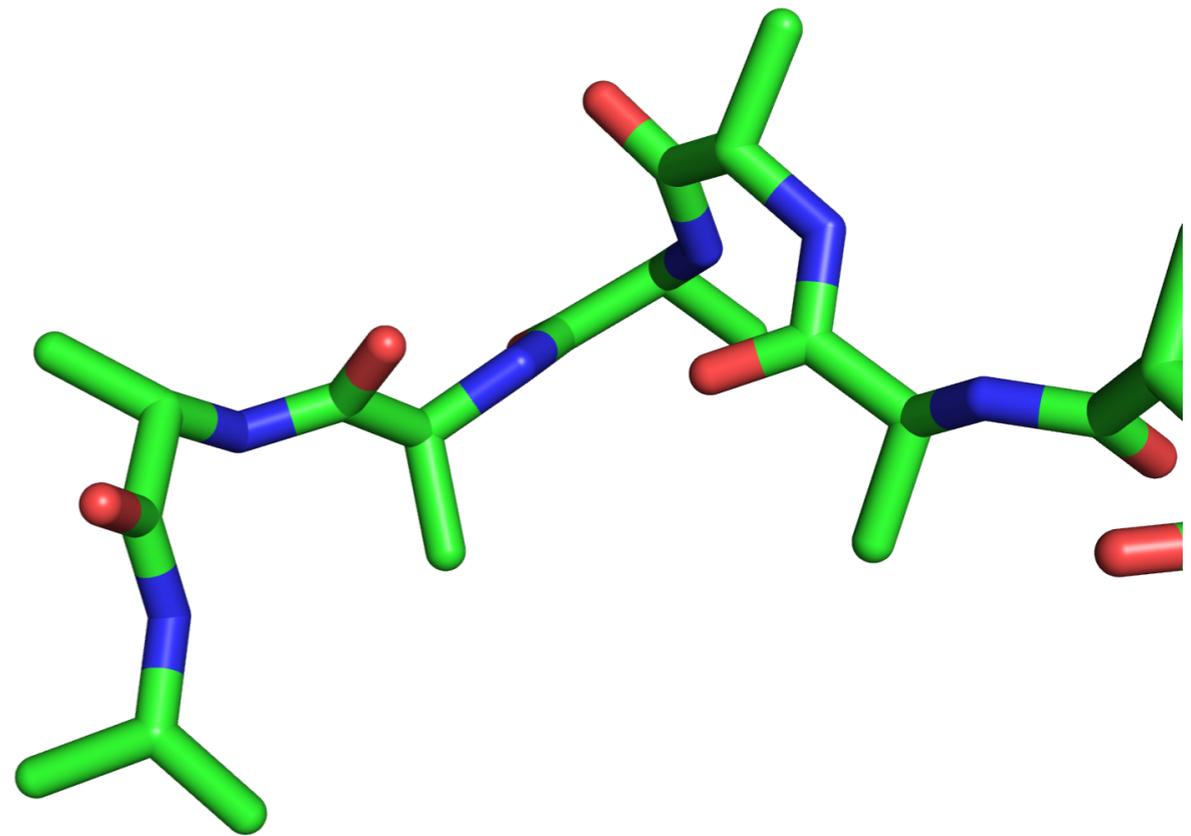
$$(\vec{x}, \vec{v}) \rightarrow (\vec{x}, \cancel{\vec{v}})$$



# Dimensionality Reduction

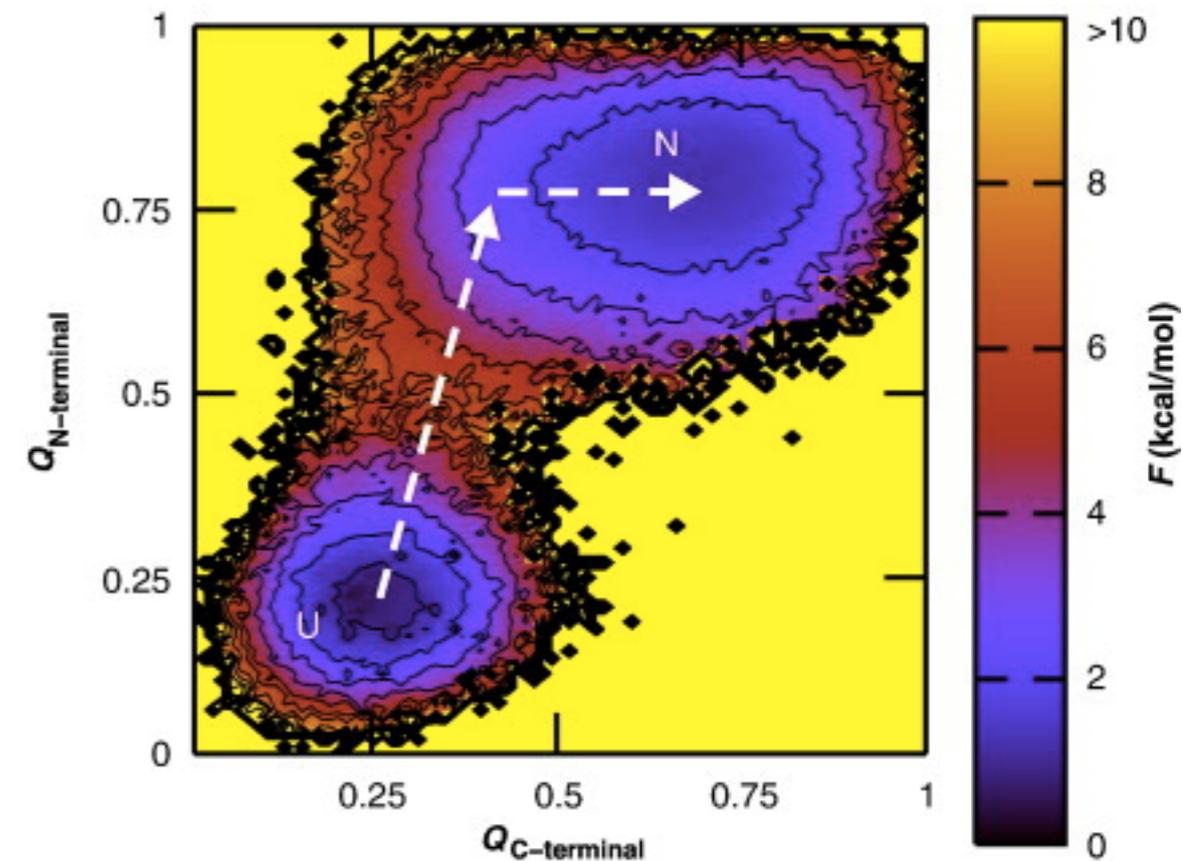
- We need to simplify the picture in order to make sense of it!
- We already do this!
  - Throw out velocities
  - Throw out solvent degrees of freedom
  - Only consider a subset of the atoms

$$(\vec{x}, \vec{v}) \rightarrow (\vec{x}, \cancel{\vec{v}})$$



# Projection-Based Analysis

- Even if we just consider a subset of all of the atoms, our dataset is usually still very high-dimensional!
- For example, a typical protein might have 500 atoms, which means we have a vector of length  $500 \times 3$  that changes in time
- The solution: turn each high-dimensional vector into one or two projections



Hills, RD Jr. and Brooks, CL III. *J. Molec. Biol.* 2008

# Common Projections

- In biomolecule simulations, several projections (reaction coordinates) are very common:
  - RMSD to a crystal pose, radius of gyration
  - Fraction of "native contacts" formed
  - Important residue - residue distances
  - DSSP assignments (secondary structure)
- In the protein folding field, many people have their "favorite" version of one of the above
- Other structural characterizations exist for non-protein systems

# Common Projections

- Root mean square deviations of atomic position (RMSD)

$$\text{RMSD}(\mathbf{X}, \mathbf{Y}) = \min_{\mathbf{R}} \sqrt{\frac{1}{n} \sum_{i=1}^n \|X_i - (\mathbf{R}\mathbf{Y})_i\|^2}$$

s.t.  $\mathbf{R}$  is a rotation matrix

- Radius of gyration

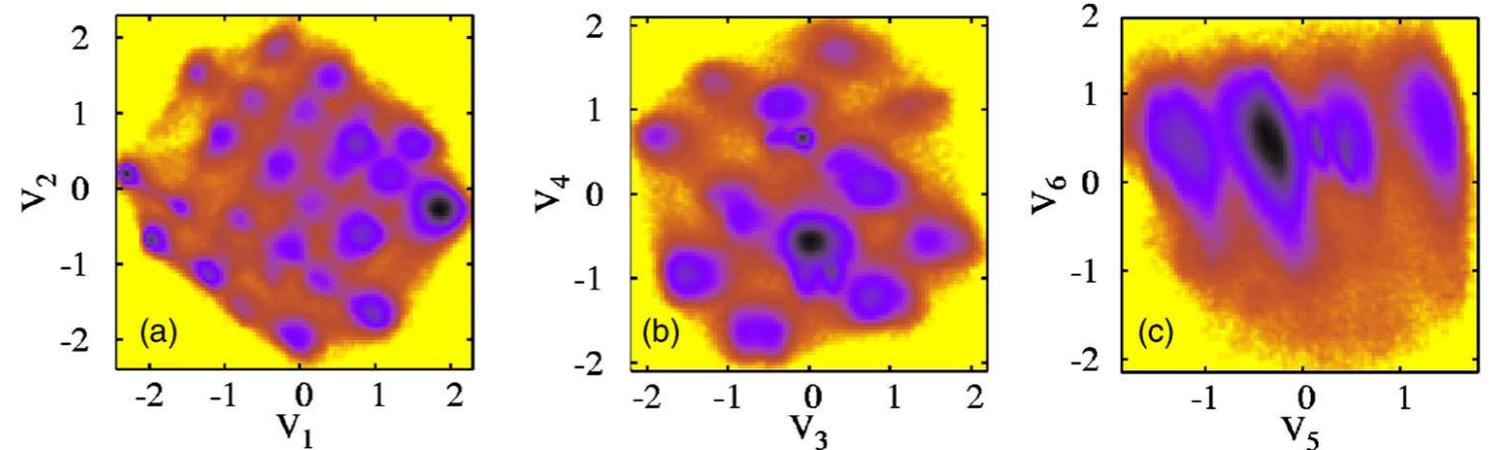
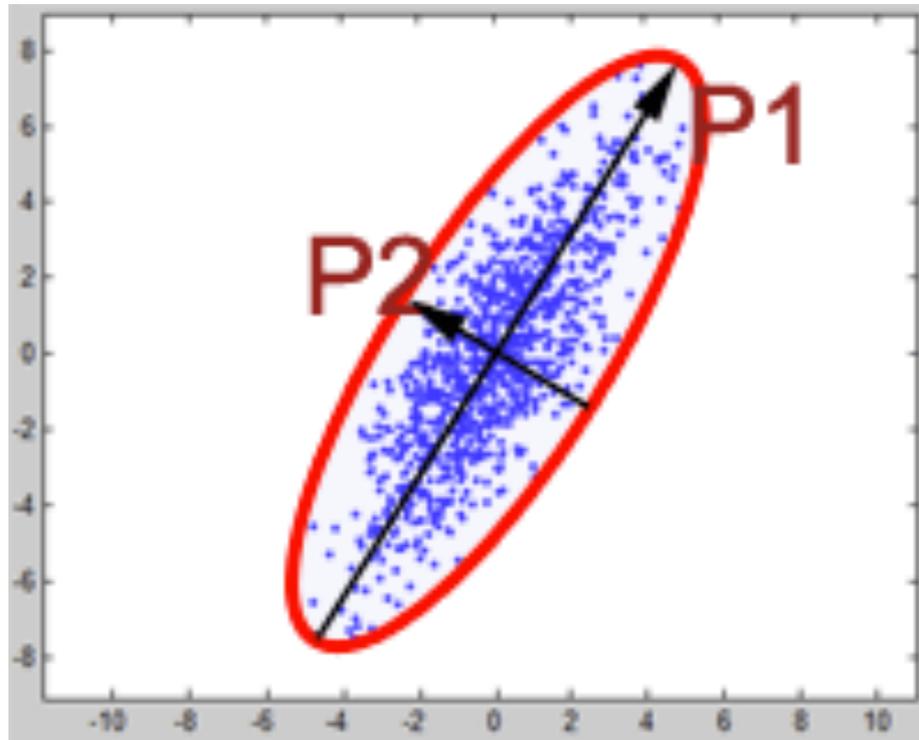
$$R_g(X) = \sqrt{\frac{1}{n} \sum_{i=1}^n (X_i - X_{mean})^2}$$

- Fraction of native contacts

$$q(\mathbf{X}) = \sqrt{\frac{1}{|\mathcal{C}|} \sum_{c \in \mathcal{C}} I_{(c \text{ formed in } \mathbf{X})}}$$

# Statistical Projections

Altis A, et al. *J. Chem. Phys.* **2008**

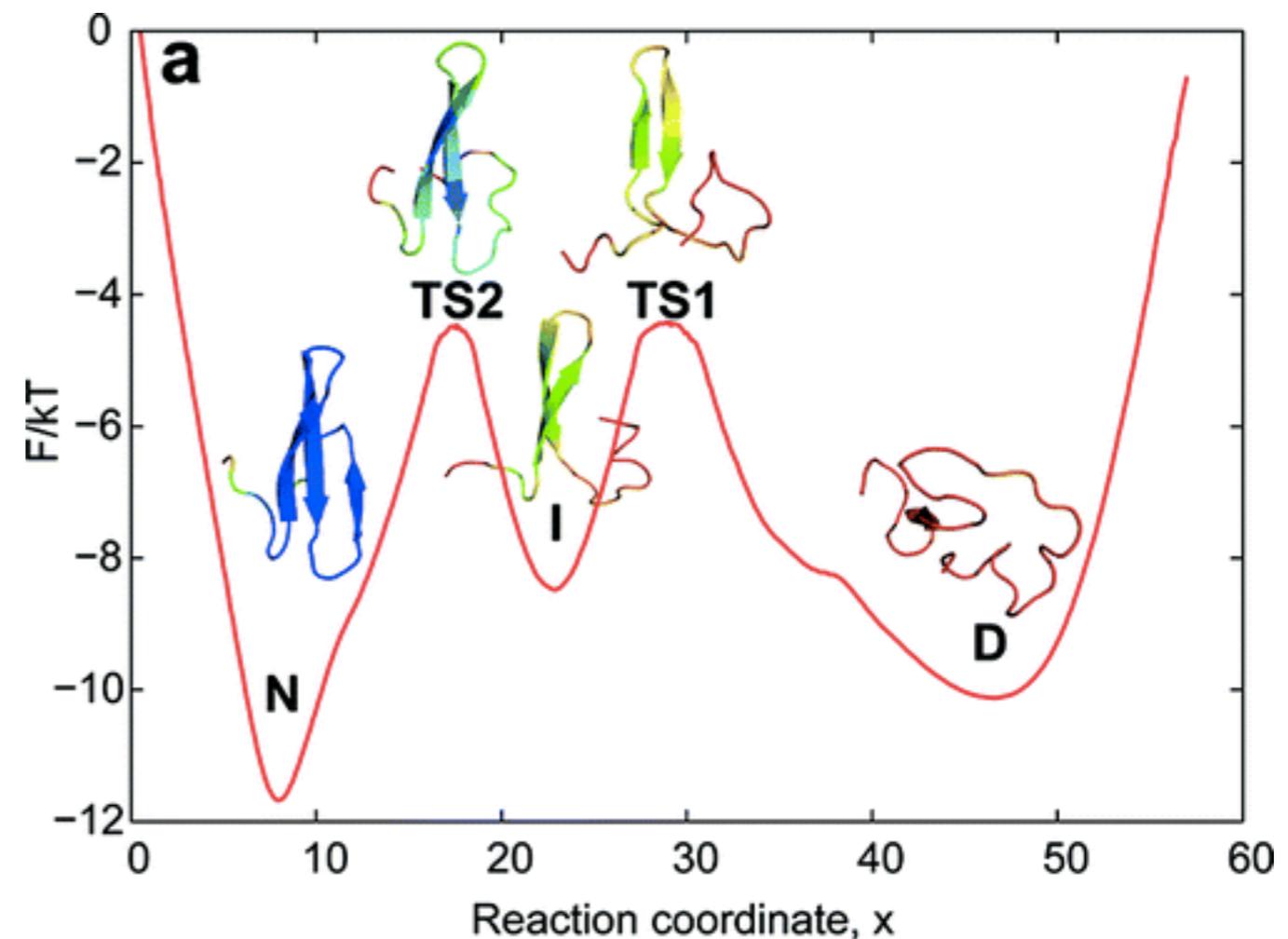


*PCA applied to protein folding simulations shows many free energy minima in the PC space*

- Another common tool is Principal Components Analysis (PCA), which looks for a projection that has maximal variance
- This is useful for exploratory analysis, but assumes that high variance is an indicator of "importance"

# Kinetic Analysis of Projections

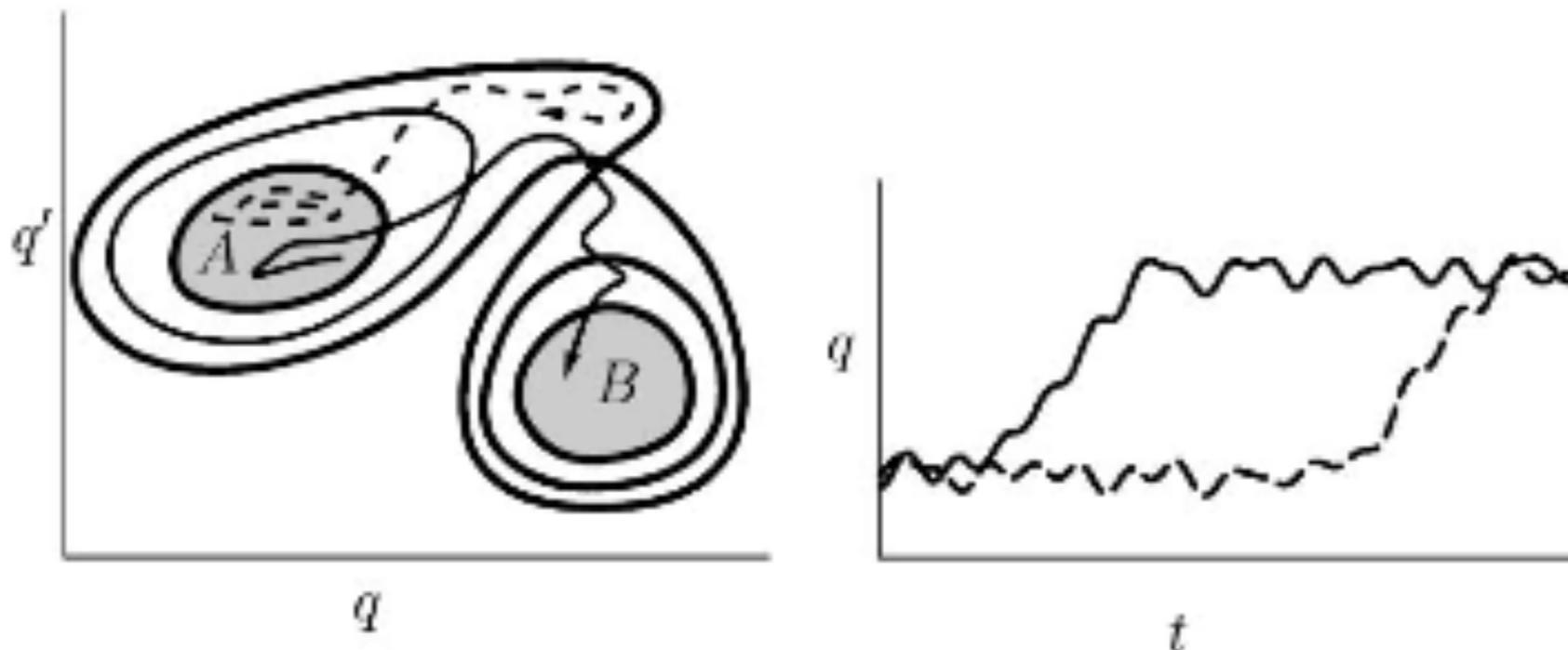
- If dynamics along the projection (reaction coordinate) are *slower* than dynamics in the orthogonal subspace, then dynamics can be modeled in the projection
- The orthogonal subspace acts like a heat bath
- But the analysis will depend on how good your reaction coordinate is



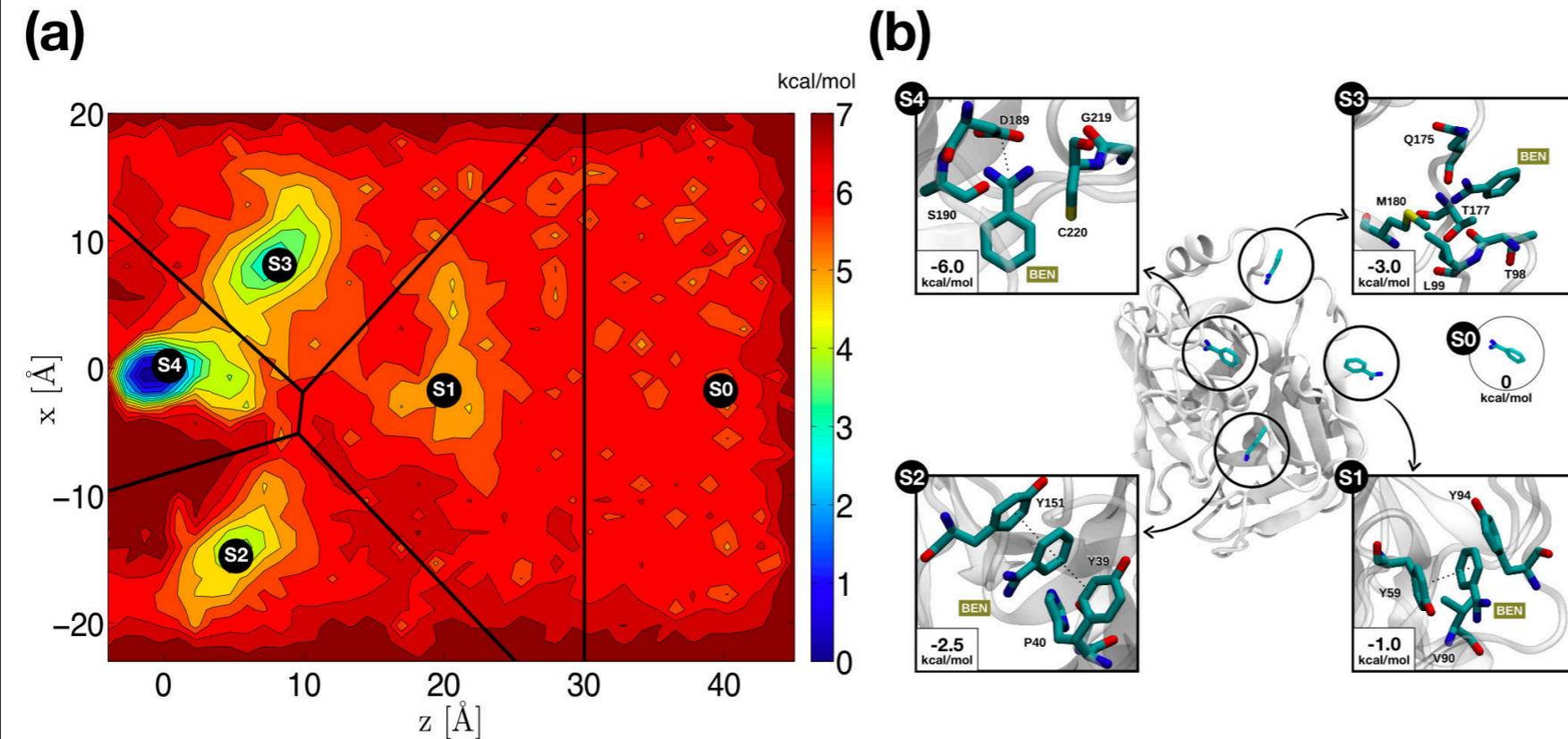
Krivov S. V. *J. Phys. Chem. B* **2011**

# Why Not Stop There?

- Projections can filter out *critical information*.
- Say, you're analyzing the potential below and asking how long it takes to go from A to B
- *If you just monitor the variable  $q$ , then you may think you've transitioned to B when in fact you haven't!*



# Motivated Projections



In protein ligand binding, a really easy projection that works well, is the location of the ligand relative to the protein

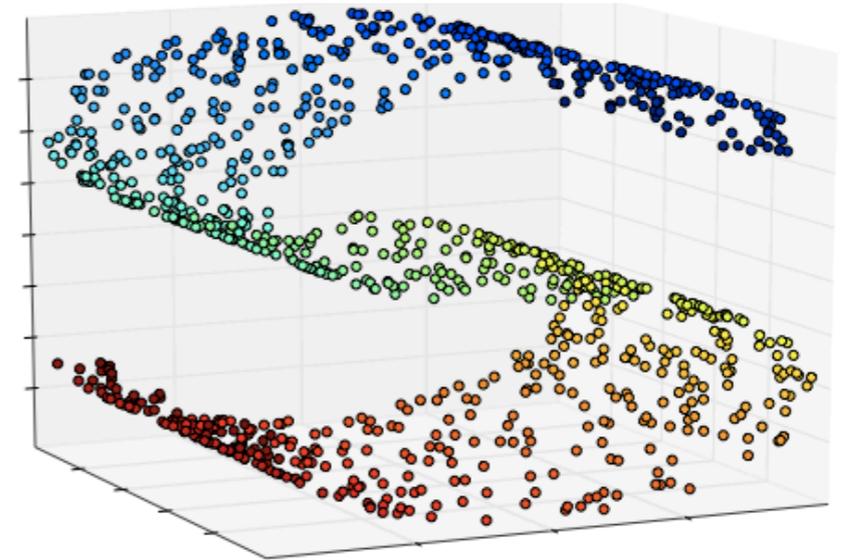
Buch, I. *et al.* *PNAS* **2011**

- In order to be confident in a projection-based method you need to know that you're picking the right thing
- In many systems, you actually already know the answer!
  - Conformational changes in kinases or well-studied enzymes
  - Protein-ligand distance

# (Machine) Learning Projections

- There are other projection based methods that attempt to pick the *correct* degrees of freedom in an automated way
- ISOMAP / Diffusion Maps (nonlinear)
- tICA (use time)
- The usefulness of these techniques will depend on the properties of your data

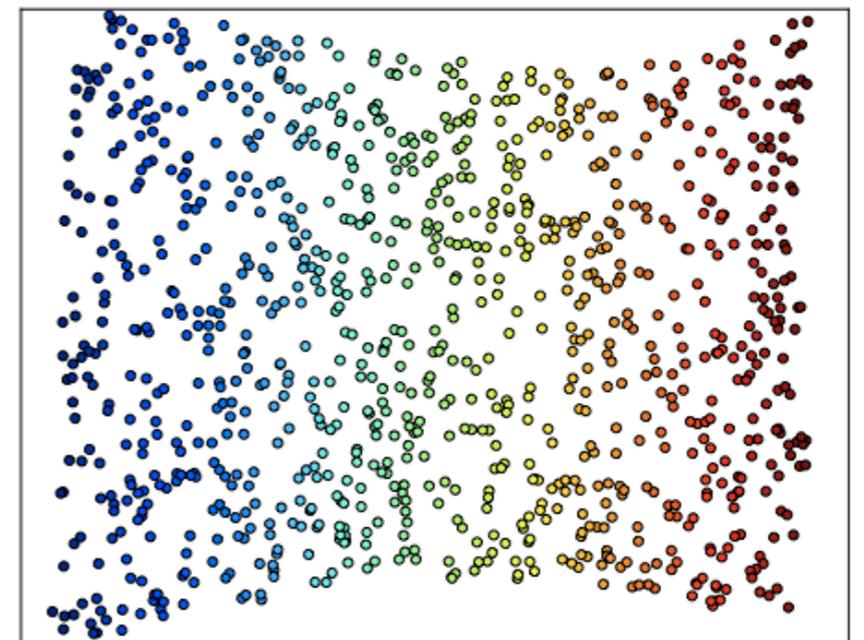
3D



machine  
learning

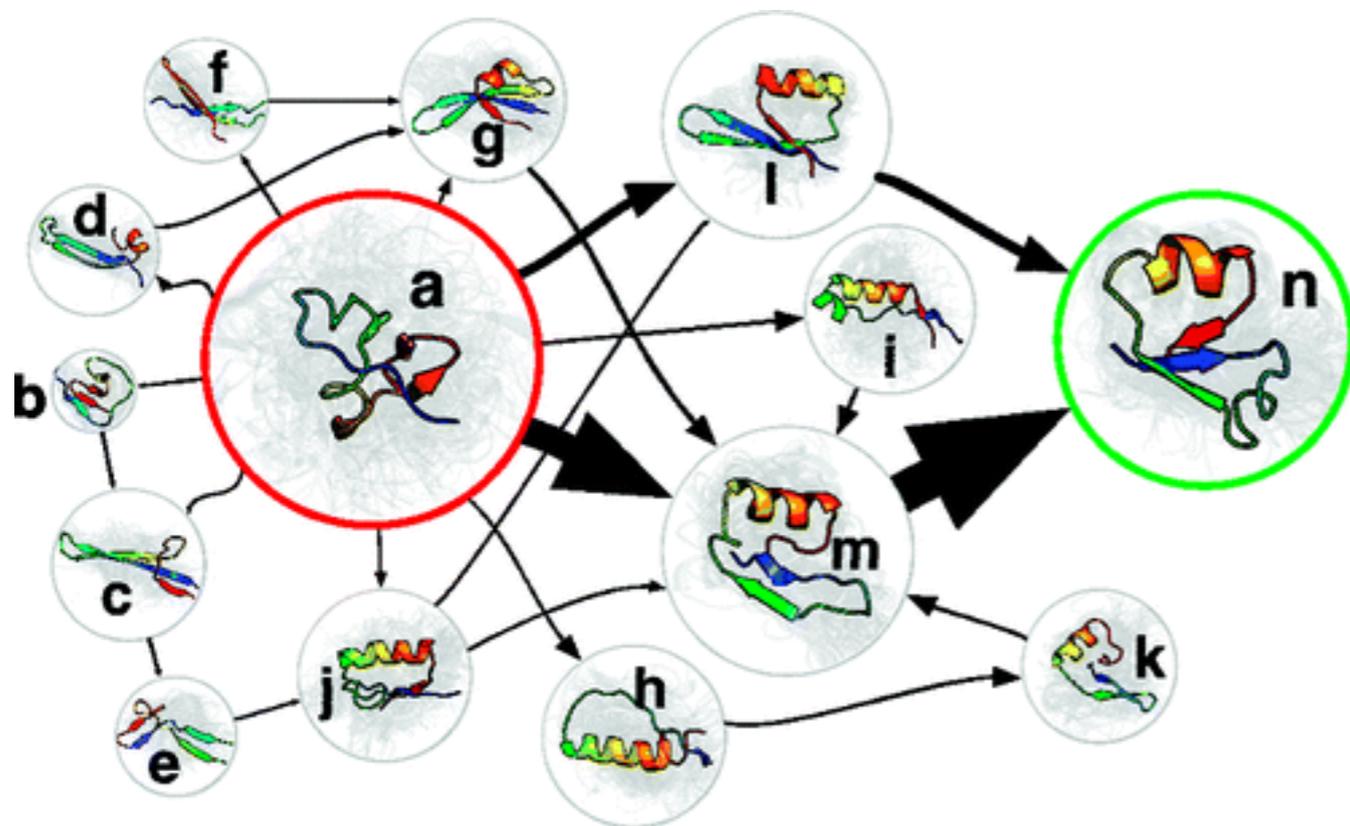


2D



# MSMs Move Beyond Projections

- Remember that projections were useful because they simplified the high-dimensional dataset into something that we could understand
- Master equations (Markov state models) approach this from a different perspective.

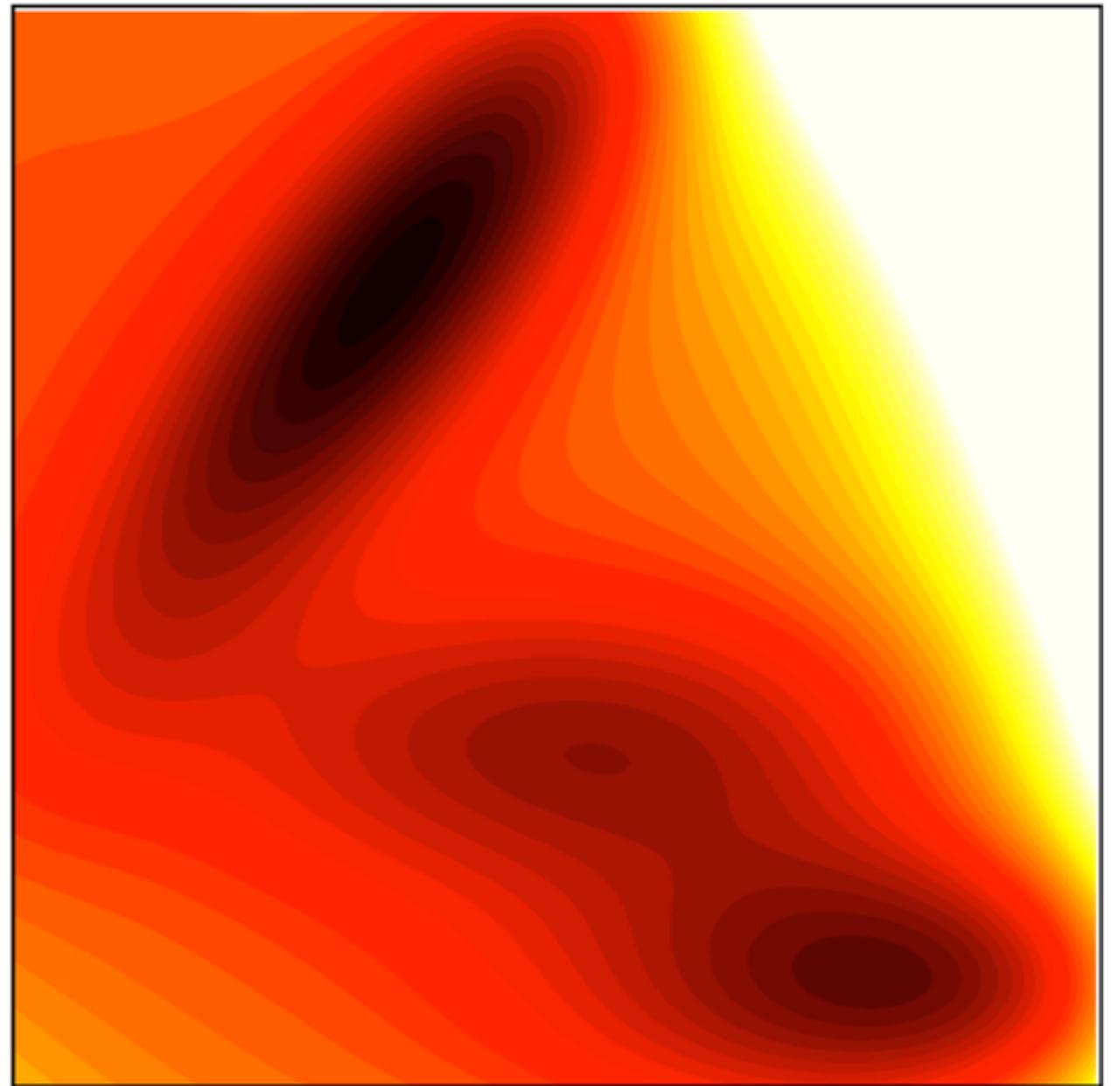


An MSM is a set of states and probabilities of transitioning between these states

***It's extendable to any simulation in which you can define the states.***

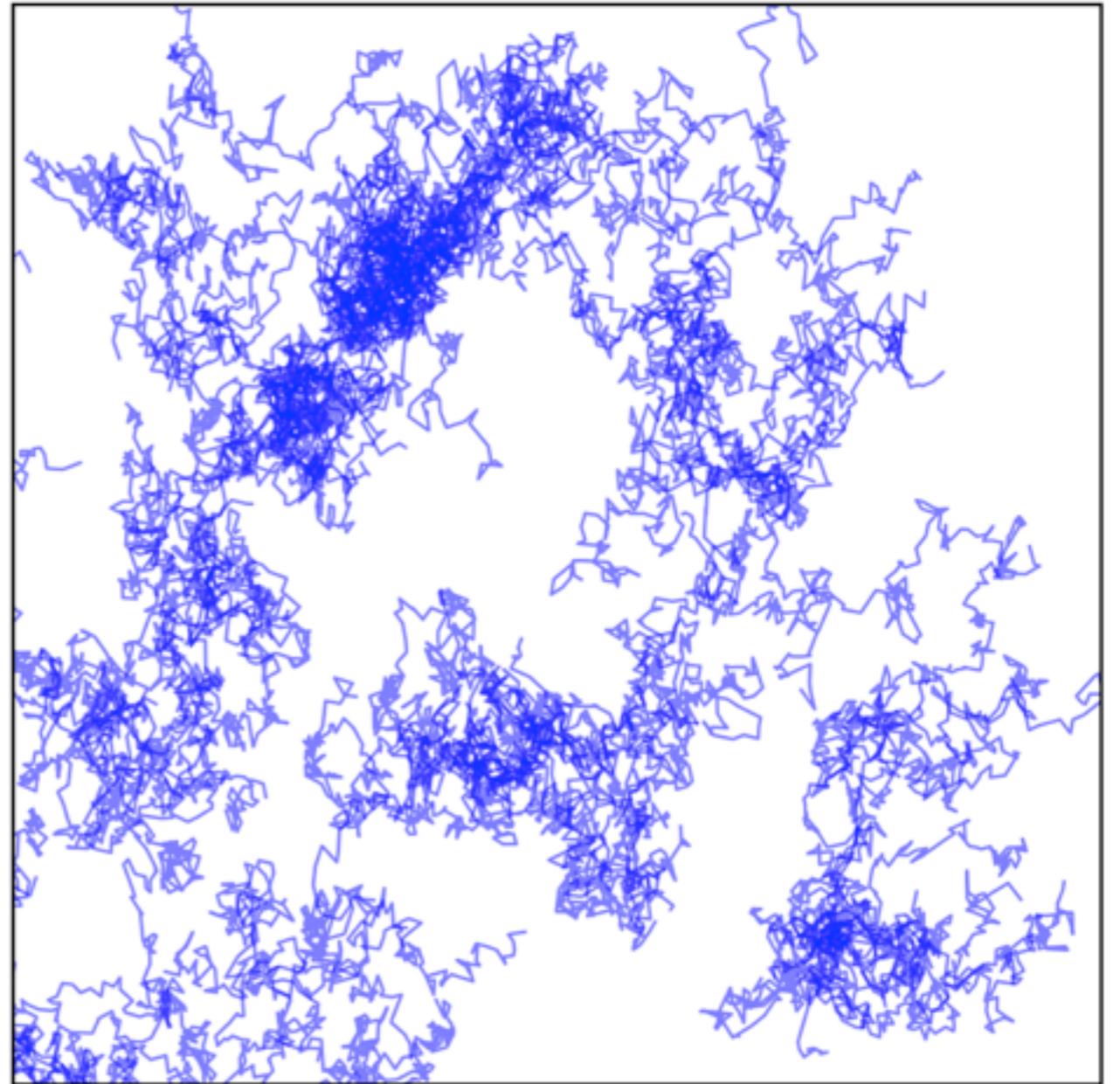
# MSM Construction

- So how do we build an MSM?
- We start with some Hamiltonian, for example the Muller potential on the right
- Sample the system with standard MD.
- The goal is to describe the thermodynamics and kinetics in terms of a set of states and rates



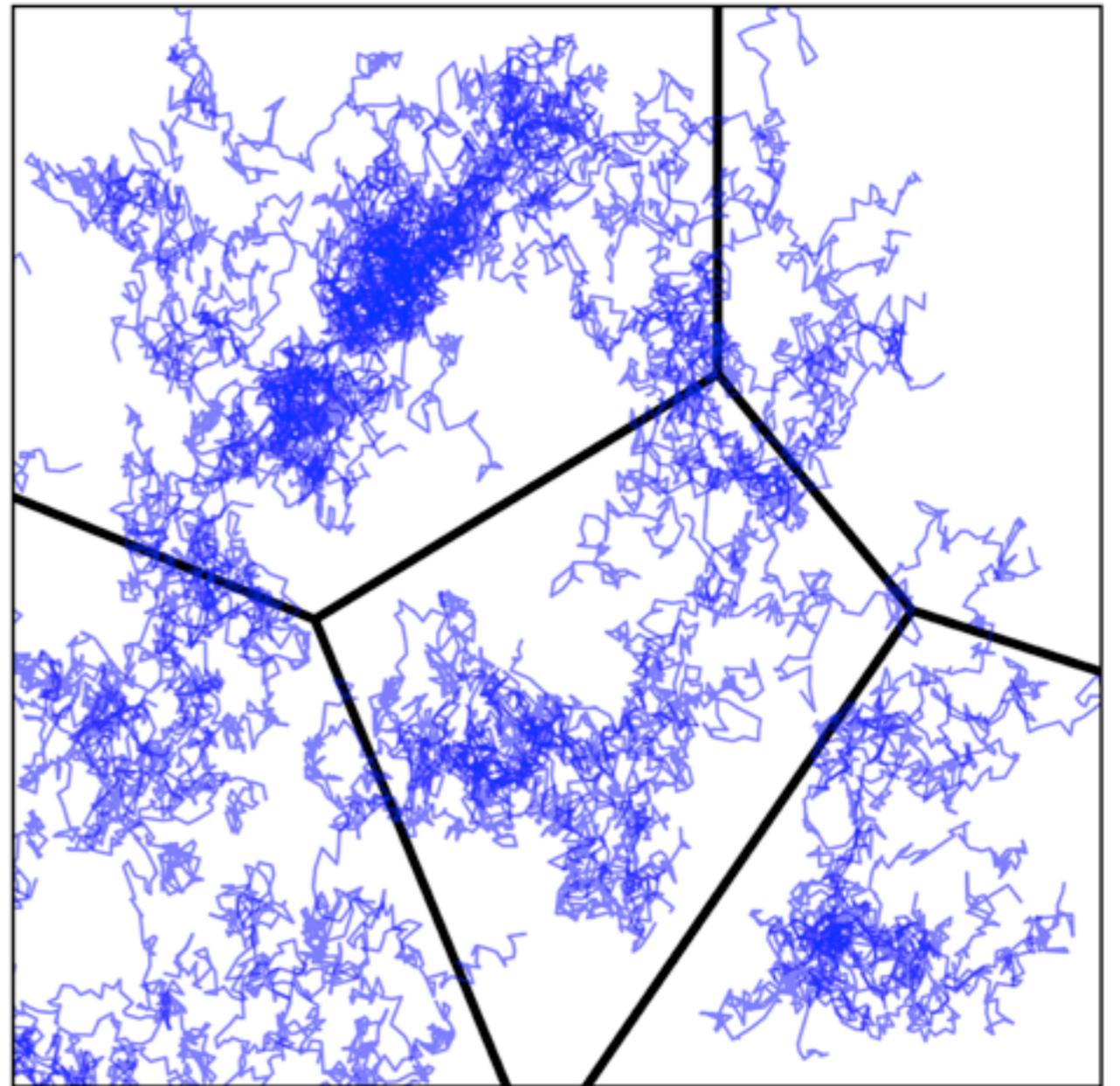
# MSM Construction

- Sampling is not easy, but can be aided by:
  - Enhanced Sampling techniques
  - Lots of cores
  - Fast hardware (GPUs, Anton)



# Markov State Models

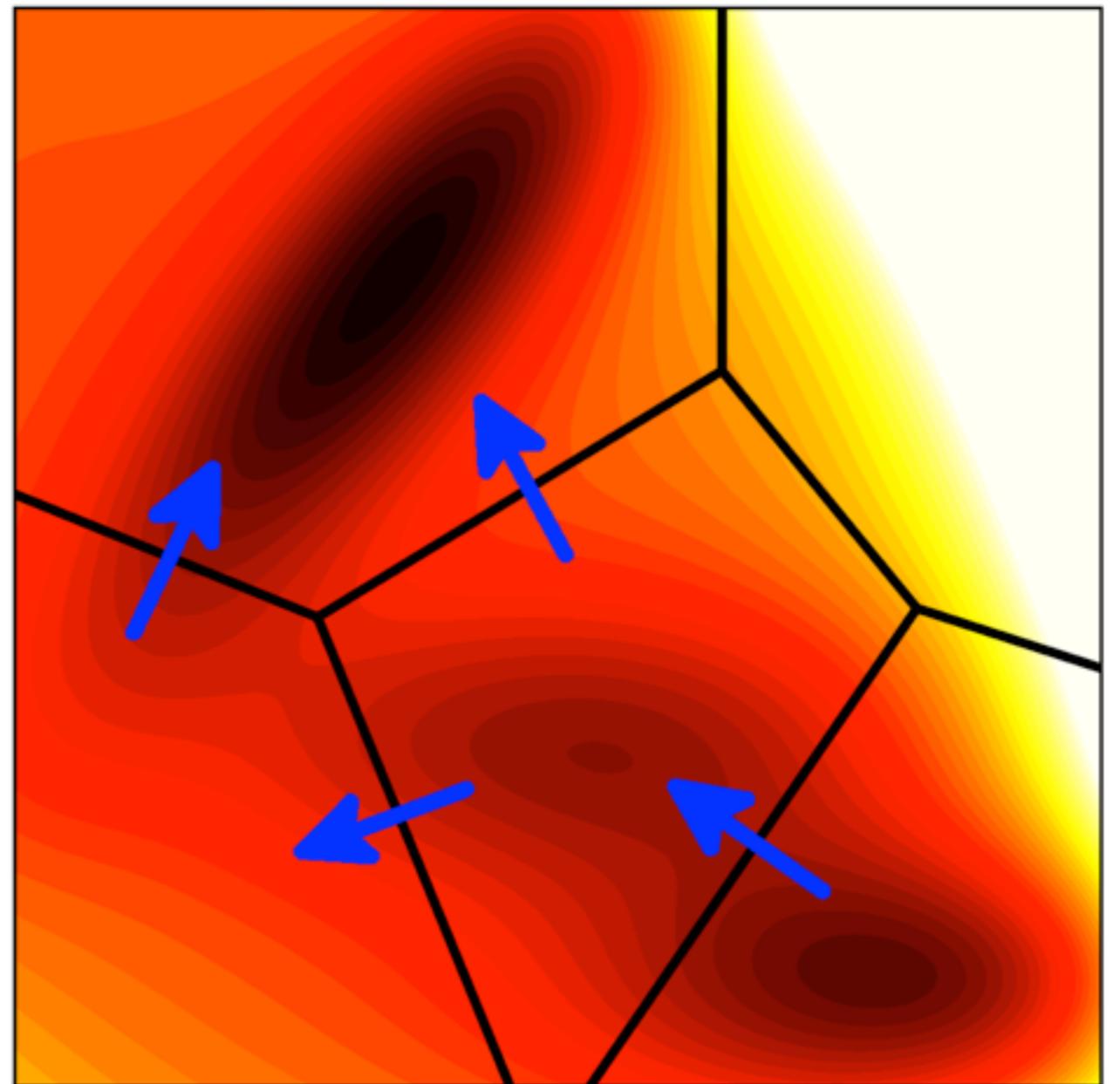
- From the sampled data, we then:
  - Define a discrete set of states
  - Calculate the rates of transferring between them
- These states should consist of points that can interconvert rapidly
- Poor state decompositions can lead to poor MSMs



# MSM Construction

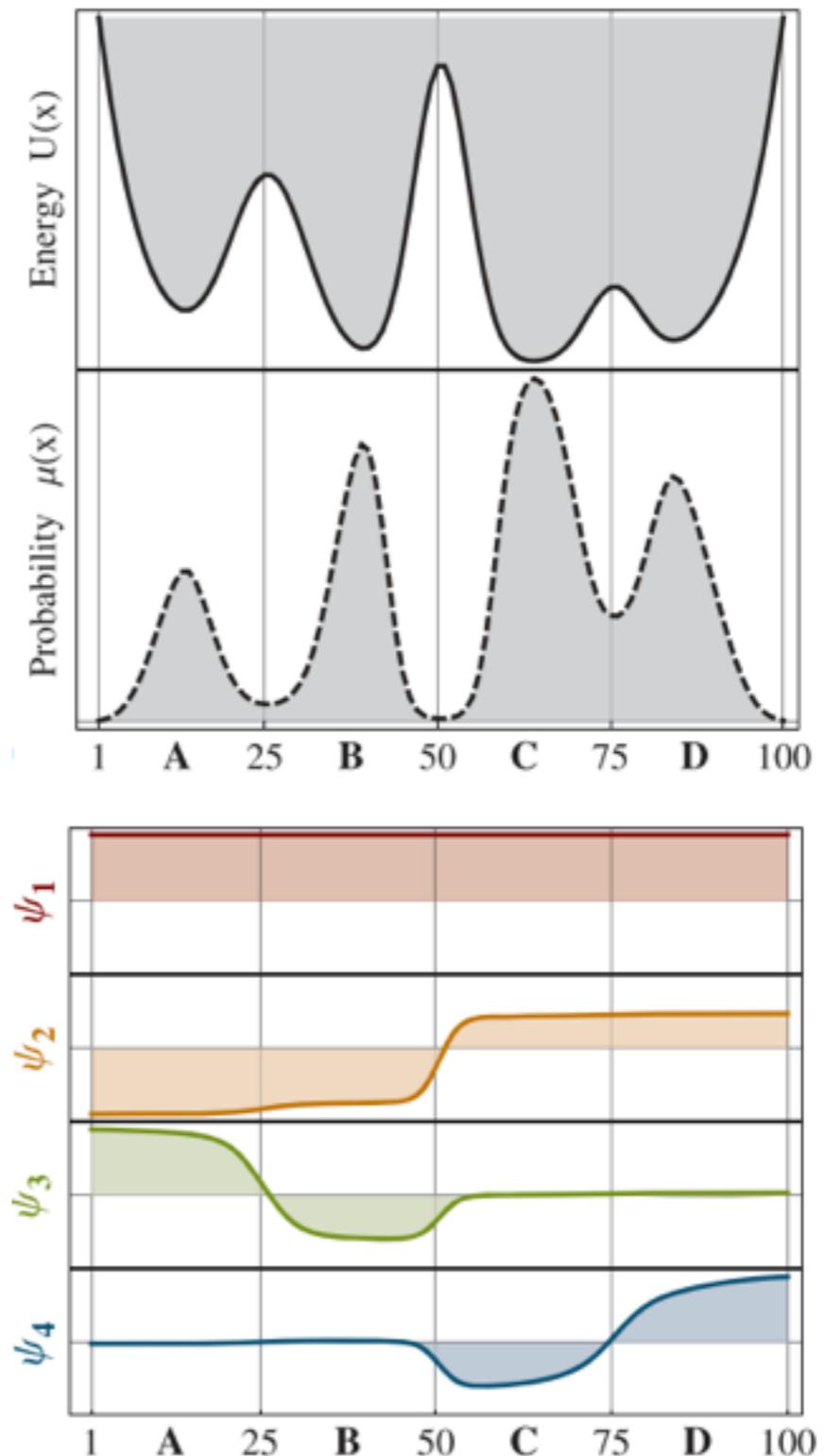
$$\vec{p}(t + \tau) = \vec{p}(t) \mathbf{T}$$

- We now have a model for the dynamics of our system
- There are many practical issues that come up in the process:
  - *How many states should we use?*
  - *How should the states be arranged spatially?*
  - *How do we validate an MSM?*



# MSM Analysis

- Now that we have an MSM, what can we do with it?
- Characterize long timescale dynamics (eigenspectrum of  $\mathbf{T}$ ).
- Find “macro-states” that are mutually kinetically separated.
- Calculate the mean first passage time between sets of states
- Quantitatively compare to / predict experimental results
- Probe mechanism



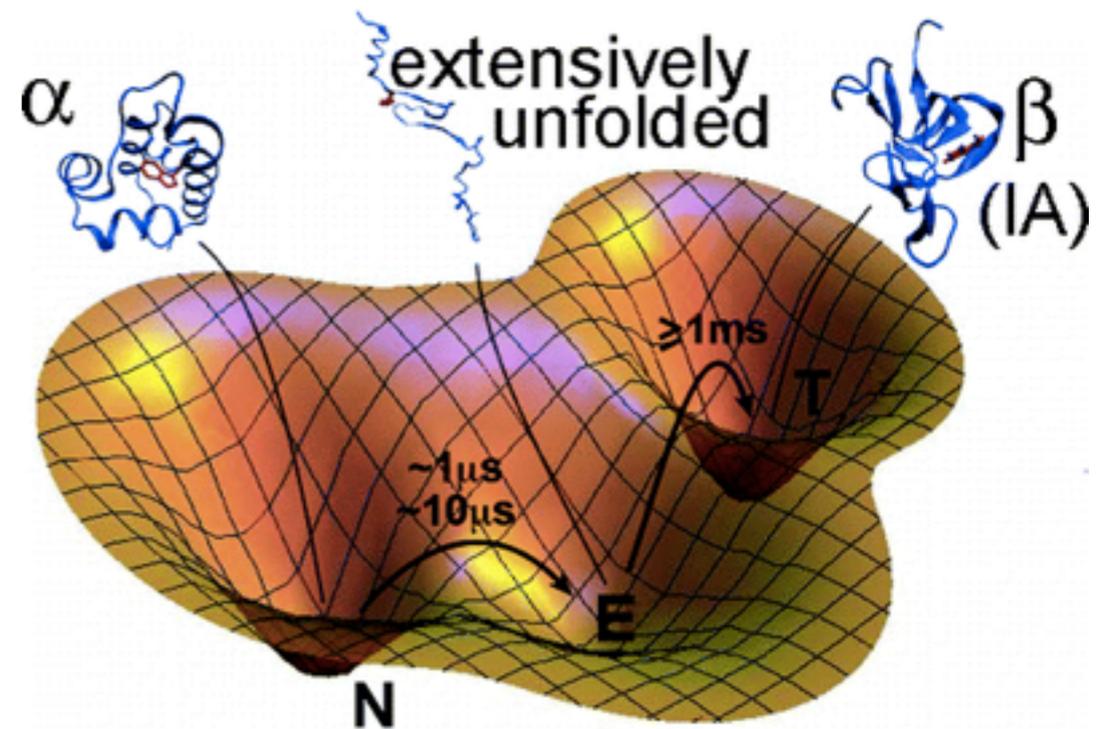
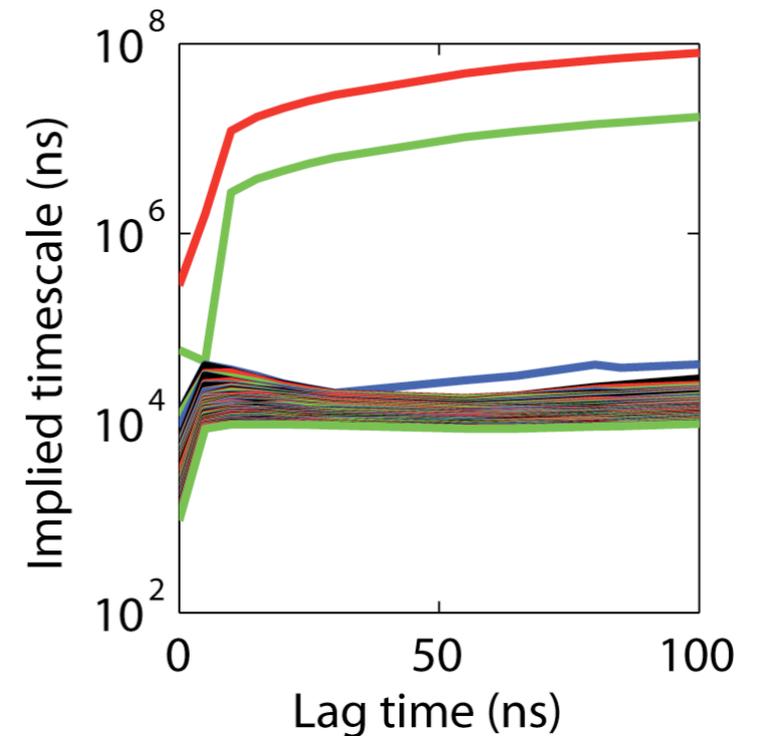
# Slowest Timescales

- The MSM's transition probability matrix can be decomposed into a sum of relaxation timescales:

$$\begin{aligned}
 p(t + n\tau)^T &= p(t)^T T^n \\
 &= \sum_{i=1}^{\infty} \lambda_i^n \langle p(t), \psi_i \rangle \phi_i \\
 &= \sum_{i=1}^{\infty} \exp\left(-\frac{n\tau}{t_i}\right) \langle p(t), \psi_i \rangle \phi_i
 \end{aligned}$$

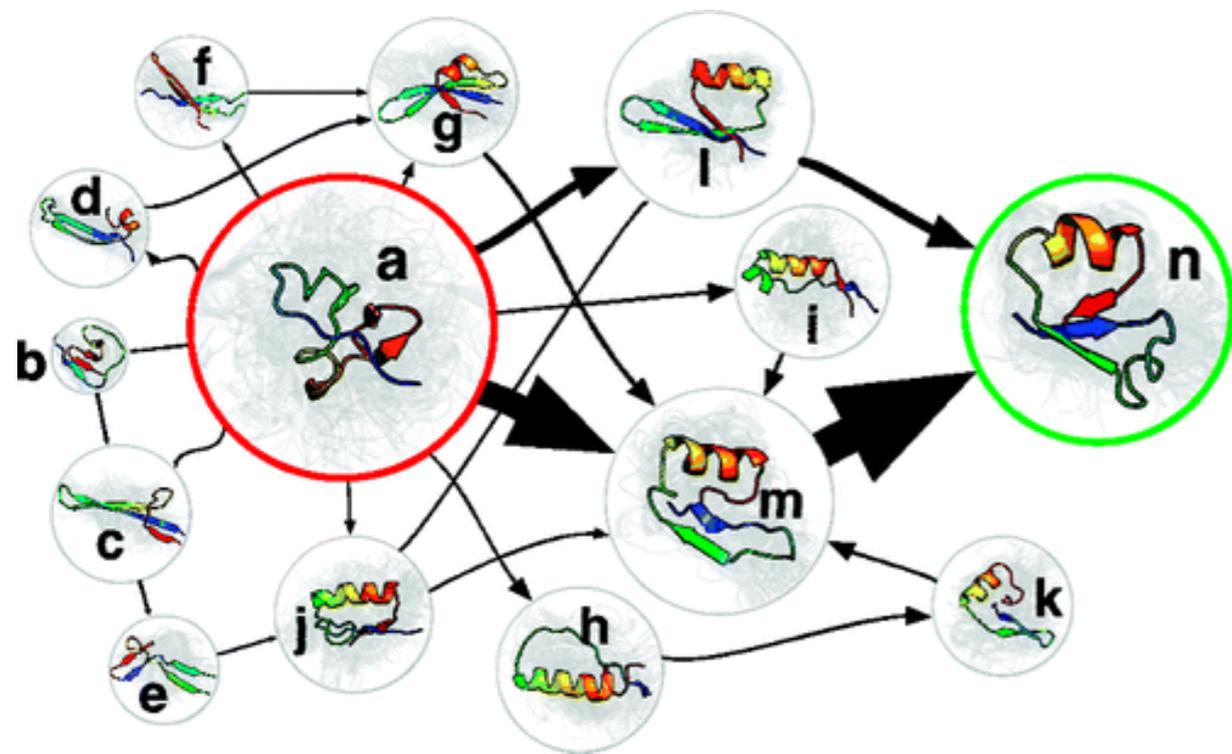
- Protein folding simulations typically have a slowest timescale corresponding to the folding transition

Bowman, G. et al. JACS 2011



Prigozhin, M. B. et al. JACS 2011

# Lump it!

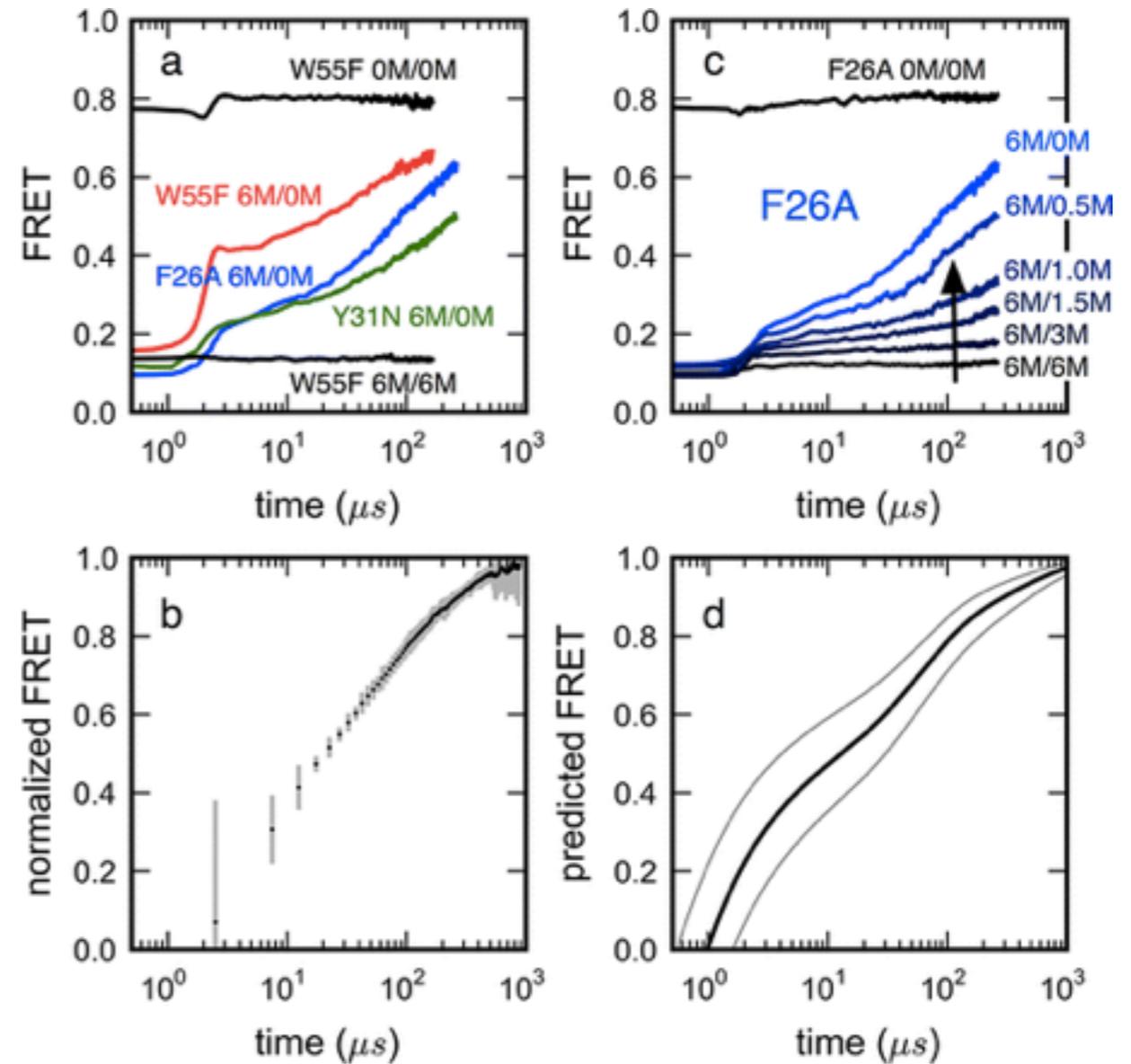


Voelz, V.A. *et al.* *JACS* **2010**, *132*, 1526-1528.

- Typical MSMs contain thousands of states
- This is simpler than what we started with, but it's still not simple...
- There are many schemes for "lumping" a given MSM such that the slowest timescales are preserved
- This can be used to build a smaller model so that you can understand the qualitative features

# Compare to Experiment

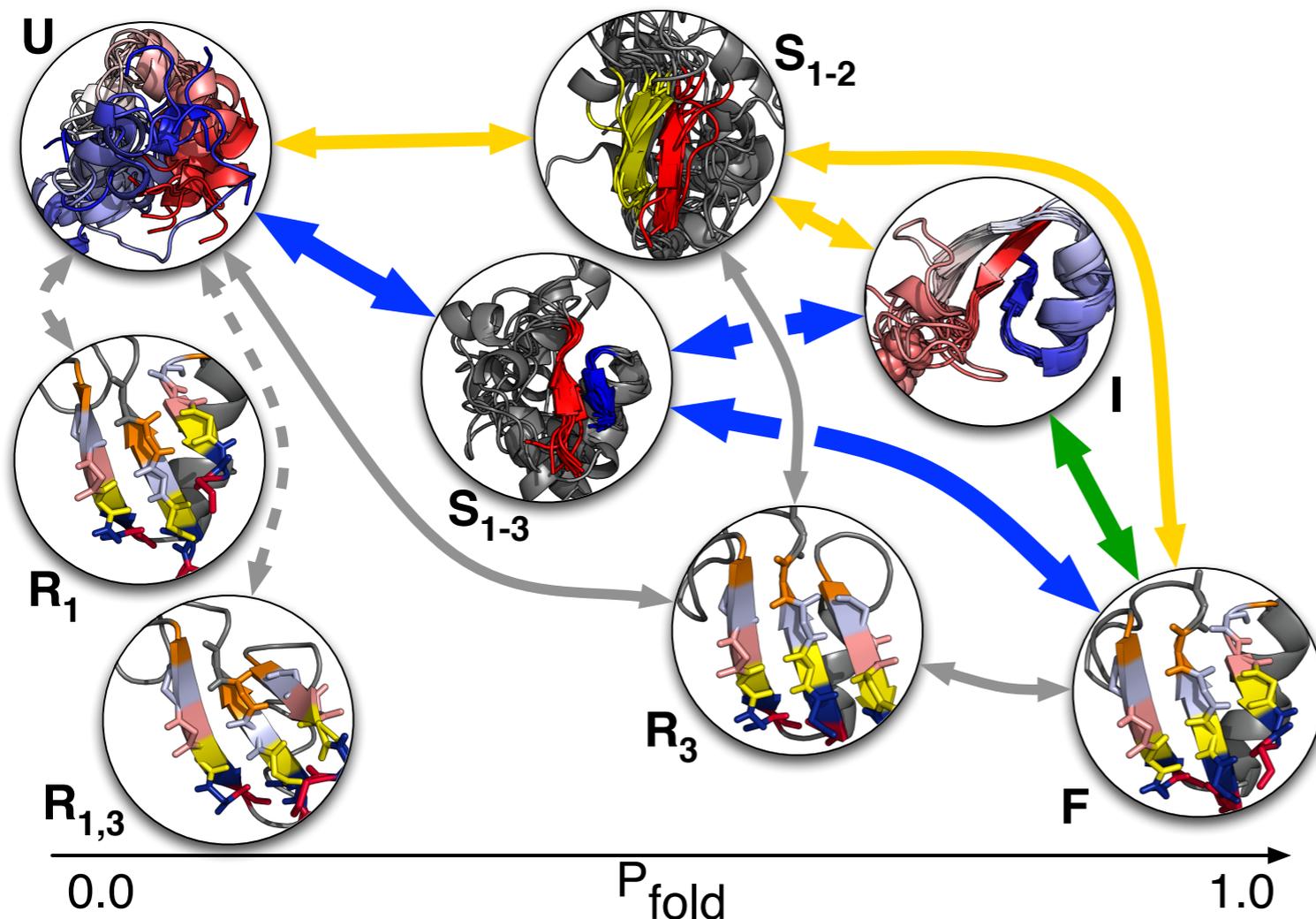
- Since we can propagate any trajectory in the MSM, it's trivial to calculate experimental observables along the way!
- We can do this in a quantitative fashion
- What you'll typically find is that one or two eigenvectors end up being prominent features in certain experiments
  - This allows you to provide a molecular interpretation of an experiment



Voelz, et al. JACS 2012

# Determine the Mechanism

- For protein folding, at least, many are interested in how a protein goes from unfolded to folded
- Within the MSM framework, you can calculate the most probable transition paths (via Transition Path Theory [TPT])



TPT often reveals many on-pathway intermediates that would be difficult to pick out while watching a movie

Schwantes, C.R. *et al.* *JCTC* **2013**